

Effect of Oxygenation during Wound Healing in Association with Root Coverage Procedures: A Pilot Clinical Trial¹Dr Manish Gupta, Department of Periodontology, Army Dental Centre(R&R), Dhaula Kuan, New Delhi²Dr T Prasanth, Department of Periodontology, Army Dental Centre(R&R), Dhaula Kuan, New Delhi³Dr Saravanan SP, Department of Periodontology, Army Dental Centre(R&R), Dhaula Kuan, New Delhi⁴Dr Ruchi Harish, Department of Periodontology, Army Dental Centre(R&R), Dhaula Kuan, New Delhi⁵Dr Manish Rathi, Department of Periodontology, Army Dental Centre(R&R), Dhaula Kuan, New Delhi**Corresponding Author:** Dr T Prasanth, Department of Periodontology, Army Dental Centre(R&R), Dhaula Kuan, New Delhi**Citation of this Article:** Dr Manish Gupta, Dr T Prasanth, Dr Saravanan SP, Dr Ruchi Harish, Dr Manish Rathi, “Effect of Oxygenation during Wound Healing in Association with Root Coverage Procedures: A Pilot Clinical Trial”, IJDSIR – July – 2026, Volume – 9, Issue – 4, P. No. 106 – 112.**Copyright:** © 2026, Dr T Prasanth, et al. This is an open access journal and article distributed under the terms of the creative common’s attribution non-commercial License. Which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given, and the new creations are licensed under the identical terms.**Type of Publication:** Original Research Article**Conflicts of Interest:** Nil**Abstract**

Background: Gingival recession occurs due to the migration of the gingiva in the apical direction, leading to compromised aesthetics and root hypersensitivity. Connective tissue graft (CTG) is still the gold standard when combined with coronally advanced flap (CAF) showing strong evidence with highest predictability. However, there are potential outcome errors associated with CAF with CTG despite having dual vascularity. The principle of oxygenation of a wound during initial stages of healing was found to enhance the vascularity of grafted site. Hence in this pilot study the effects of topical application of active oxygen oral gel post root coverage surgery was assessed.

Aim & Objectives: The aim of our study was to evaluate the effect of oxygenation therapy during wound healing

after CAF with CTG using clinical parameters and with immunological marker Interleukin -1 beta (IL-1 β).

Materials and methods: Individuals aged between 16 to 60 years in test group with Cairo’s RT 2 recession, were included They were randomly assigned to test group with topical application of oxygen gel. GCF and clinical parameters were calculated at baseline (T0), One week(T1) and one month post-operatively (T2) to quantify GCF IL-1 β on pre & post active oxygen gel application.

Results: The quantitative levels of IL-1 β in test group at T0 (80.4 pg/ml) T1 (97.4 pg/ml) and at T2 (58.8 pg/ml) and ANOVA test yielded statistically significant difference in IL-1 β level with topical application of oxygen gel which exhibited in IL-1 β count, which sustained for a period of one month (p= 0.03).

Conclusion: The study revealed that topical oxygen gel application has significant benefit in wound healing associated with root coverage procedures, Notably, this study observed a significant reduction in IL -1 β in the immediate post operative period favouring optimal healing outcome.

Keywords: Cavities, Gum Tissue, Malocclusion, Oxygenation Gel

Introduction

Gingival recession is indeed a common oral condition characterized by the exposure of the root surface of a tooth due to the migration of the gingiva in the apical direction. This migration can lead to several potential issues for the patient, including an increased risk of dental caries (tooth decay), erosion of the root surface, increase in tooth sensitivity, and compromised esthetics.^{1,2.}

The etiological factors ^{3,4} contributing to gingival recession are diverse and can include:

Inadequate Restorations: Poorly restored dental restorations or fillings can contribute to gingival recession.

Malpositioned Teeth: Teeth that are not properly aligned can exert pressure on the surrounding gingival tissue, leading to recession.

Cavities: Untreated dental cavities can cause localized destruction of the tooth and surrounding gum tissue, potentially resulting in recession.

Periodontal Disease: Gum disease, such as gingivitis or periodontitis, is a significant cause of gingival recession. The inflammation and infection associated with these conditions can lead to tissue destruction and recession.

Labial Frenulum: In some cases, an inadequate insertion of the labial frenulum (the tissue connecting the lip to the gums) can contribute to gingival recession.

Thin Tissue Phenotype: Individuals with a thin tissue phenotype are more prone to gingival recession due to reduced tissue thickness and resilience.

The classification¹⁰ of gingival recession has evolved over time to provide a more precise understanding of the condition. The new classification, based on the measurement of interdental clinical insertion, categorizes gingival recessions into three types:

Type 1 Recession (RT1): Gingival recession without loss of interproximal insertion.

Type 2 Recession (RT2): Gingival recession with loss of interproximal insertion, where the measurement of the cemento-enamel junction is less than or equal to the loss of buccal insertion.

Type 3 Recession (RT3): Gingival recession with loss of interproximal insertion, where the measurement of the cemento-enamel junction is greater than the loss of oral insertion.

This classification system helps clinicians in diagnosing and planning treatment for gingival recession by considering both the extent of recession and the degree of attachment loss⁵. Effective management of gingival recession often involves addressing the underlying causes, such as periodontal disease or malocclusion, and employing appropriate surgical techniques, such as gingival grafting, to restore tissue health and aesthetics.

Connective tissue graft (CTG) is still the gold standard when combined with coronally advanced flap (CAF) showing strong evidence with highest predictability⁶. However there are potential outcome errors associated with CAF with CTG despite having dual vascularity. The principle of oxygenation of a wound during initial stages of healing was found to enhance the vascularity of grafted site. Hence in this pilot study the effects of topical application of active oxygen oral gel post root coverage surgery is assessed.

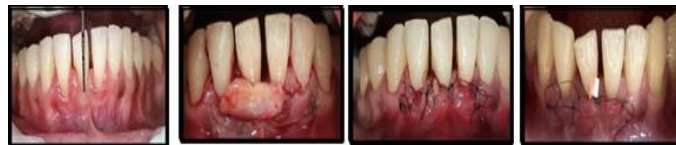
Materials and Method

A single blind randomized controlled, prospective clinical study was conducted for three months with 10 participants assigned to test and control group using block randomisation method. Patients with Cairo's RT 2 recession were considered in the study. Both males and females in the age group of 25-50 years who attended the outpatient department of Periodontics, were included in the study. The Institutional Ethical clearance was obtained and it was monitored in accordance with Declaration of Helsinki and Good Clinical Practice (ICH-GCP) guidelines. Smokers and alcoholics, pregnant and lactating women, patients treated with antibiotics in the past 6 months, participants allergic to lactose and fermented milk products and to other drugs, patients with systemic illness and HIV infection/AIDS were excluded from the study. Prior to initiating the study, the patients were informed of the purpose and design of the study and were requested to sign an informed consent.

After the connective tissue graft (CTG) was harvested on the recipient site, for the test group an oxygenation gel was applied at the recipient site for seven days after the surgery. Gingival crevicular fluid (GCF) samples were collected from all participants at three different time points: T0 (before the surgery), T1 (after 7 days of surgery), and T3 (after one month of surgery). This study design allows for the assessment of the effects of the oxygenation gel on the healing process and GCF Interleukin 1- β levels over time.

GCF Interleukin 1- β levels was assessed by paper point method (Quantitative Assay using ELISA). GCF sample was collected by gently inserting sterile paper point into gingival sulcus for 20 sec and placed in Eppendorf tube containing Phosphate Buffer Solution and stored at -80 degree until needed.

All the samples were subjected to analysis of IL 1 beta. Diaclone Human IL-1 β ELISA Kit was used for analysis of GCF Interleukin 1- β levels.



Recession Coverage procedure for Test and Control Group



Oxygenation gel application for Test Group



IL-1 β Kit



GCF Solution embedded in well



Spectrophotometer

Results

The study design allows for the assessment of the effects of the oxygenation gel on the healing process and GCF Interleukin 1- β levels over time. Analysing GCF samples

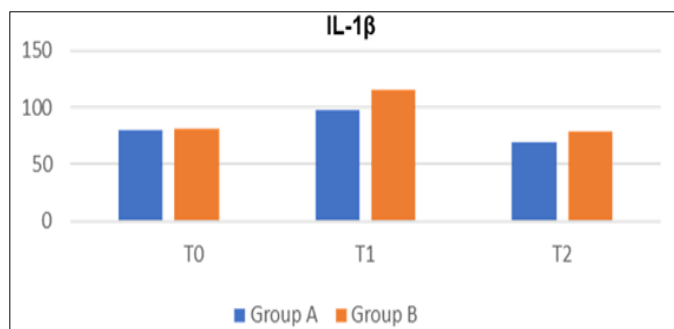
at these time points can provide valuable insights into the inflammatory response, tissue healing, and potential effects of the oxygenation gel by quantify GCF IL-1 β on pre & post active oxygen gel application.

The patients included the study comprised of 06 male and 04 female patients aged between 28-60 year. The study included two groups, where group A subjects were treated with oxygenation gel application after root coverage procedure and the group B, subjects treated with root coverage procedure only and The GCF samples were taken for both the groups at baseline, 1 week and 1 month The statistical analysis was carried out using ANNOVA.

The quantitative levels of GCF Interleukin 1- β levels Group A at T₀ T₁ and at T₂ was 80.4pg/ml, 97.4pg/ml & 58.8pg/ml respectively and in Group B it was found to be 81.2pg/ml (T₀), 115.2pg/ml(T₁) and 78.5pg/ml (T₃). ANOVA test yielded statistically significant results in the root coverage procedure with oxygenation gel application. ($p=0.03$).

	Group A	Group B
T ₀	80.4 pg/ml	81.2 pg/ml
T ₁	97.4 pg/ml	115.2 pg/ml
T ₂	58.8 pg/ml	78.5 pg/ml

Quantitative analysis of IL-1 β Level at baseline, 1 week and after 1 month



Discussion

Gingival recession can lead to both physical discomfort and aesthetic concerns for patients. Root coverage

procedures aim to address these issues by increasing the keratinized gingiva band⁶, which helps protect the underlying tissues, improving aesthetics and functionality. Additionally, eliminating dentin hypersensitivity² provides relief from discomfort associated with exposed dentin surfaces, enhancing patient comfort. Moreover, by minimizing the accumulation of plaque^{7,8} root coverage procedures contribute to the prevention of periodontal diseases and support overall oral health. Therefore, an effective root coverage approach should encompass these objectives to ensure comprehensive care for patients with gingival recession.

Wound healing is a complex process, which it has oxygen as a prerequisite in almost every step. Topical therapies that release O₂ molecules are great allies in reparative processes, favouring increased cell metabolism, collagen synthesis, antibacterial activity, release of growth factors and angiogenesis⁹.

When oxygenation gel components come into contact with moist surfaces such as mucosa, saliva, and blood, they initiate a decomposition reaction, releasing oxygen in the form of hydrogen peroxide into the surrounding medium. At low concentrations, this active oxygen exhibits antibacterial properties and can promote angiogenesis, increased collagen synthesis, cellular metabolism, and the release of growth factors. These effects contribute to improving the overall oxygen transport capacity, facilitating tissue repair¹⁰.

Studies, such as the one by Imano et al. (2019), have demonstrated the potential of oxygenation gel in aiding the healing and formation of keratinized tissue. In procedures like free connective tissue grafting, satisfactory healing outcomes were observed, likely due to the gel's angiogenic stimulus and antibacterial effects.

The immune response to wound healing involves the release of various cytokines, including inflammatory cytokines like IL-1 β , regulatory cytokines like IL-10, and chemokines like IL-8.¹¹ Assessing the effects of oxygenation gel on the healing process and levels of GCF interleukin-1 β (IL-1 β) over time provides valuable insights into the inflammatory response, tissue healing, and potential impacts of the gel. Quantifying GCF IL-1 β levels before and after active oxygen gel application offers a means to evaluate its efficacy in modulating inflammation and supporting tissue repair processes.

Overall, the use of oxygenation gel in wound healing holds promise for enhancing the reparative process and improving outcomes in root coverage procedures.

Conclusion

The study revealed that topical oxygen gel application has significant benefit in wound healing associated with root coverage procedures. Notably, this study observed a significant reduction in IL -1 β in the immediate post operative period favoring optimal healing outcome. New clinical trials and must be carried out to observe the effects of oxygen therapy on the technique of free connective tissue grafting.

References

1. Zucchelli, G., & De Sanctis, M. (2000). Treatment of multiple recession-type defects in patients with esthetic demands. *J Periodontol.* 71(9), 1506-15
2. Eisenbud, D. E. (2012) Oxygen in wound healing: nutrient, antibiotic, signaling molecule, and therapeutic agent. *Clin Plast Surg.* 39(3):293-310. 10.1016/j.cps.2012.05.001
3. Yadav, A. P., Kulloli, A., Shetty, S., Ligade, S. S., Martande, S. S., & Gholkar, M. J. (2018). Sub-epithelial connective tissue graft for the management of Miller's class I and class II isolated gingival recession defect: A systematic review of the factors influencing the outcome. *J Investig Clin Dent.* 9(3):e12325. 10.1002/JPER.16-0671
4. Storrer, C. L. M., Muller, L. L., Pissai, J. F., Andrade, C. F., Trevisani, C. R. T., & Deliberador, T. M. (2019) Treatment of Miller Class I Gingival Recession with Using Non pedicle Adipose Tissue after Bichectomy Surgical Technique: A Case Report. *Case Rep Dent:* 1049453. 10.1155/2019/1049453
5. Cortellini, P., & Bissada, N. F. (2018). Mucogingival conditions in the natural dentition: Narrative review, case definitions, and diagnostic considerations. *J Periodontol.*; 89 1:204-213. 10.1002/JPER.16-0671
6. Yoshino, H., Hasuiki, A., Sanjo, N., Daisuke, S., Tatesuya, K., Hidekazu, N., et al. (2020). CO2 Laser De-epithelization Technique for Subepithelial Connective Tissue Graft: A Study of 21 Recessions. *In Vivo.* 34(2):869-875. 10.21873/invivo.11851
7. Rasperini, G., Rocuzzo, M., Francetti, L., Acunzo, R., Consonni, D., & Silvestri, M. (2011). Subepithelial connective tissue graft for treatment of gingival recessions with and without enamel matrix derivative: a multicenter, randomized controlled clinical trial. *Int J Periodontics Restorative Dent,* 31(2), 133-139.
8. Zucchelli, G., Gori, G., Mele, M., Stefanini, M., Mazzotti, C., Marzadori, M., Montebugnoli, L., & De Sanctis, M. (2011). Non-carious cervical lesions associated with gingival recessions: a decision-making progress. *J Periodontol.* 82(12), 1713-1724
9. Gamonal J, Acevedo A, Bascones A, Jorge O, Silva A. Levels of interleukin-1 β , -8, and -10 and RANTES in gingival crevicular fluid and cell populations in adult periodontitis patients and the effect of periodontal treatment. *Journal of periodontology.* 2000 Oct;71(10):1535-45.

10. Ngeow WC, Tan CC, Goh YC, Deliberador TM, Cheah CW. A Narrative Review on Means to Promote Oxygenation and Angiogenesis in Oral Wound Healing. Bioengineering.2022 Nov 2;9(11):636
11. Zucchelli, G., Testori, T., & De Sanctis, M. (2006). Clinical and anatomical factors limiting treatment outcomes of gingival recession: a new method to predetermine the line of root coverage. J Periodontol. 77(4), 714-721
12. Han, J. S., John, V., Blanchard, S. B., Kowolik, M. J., & Eckert, G. J. (2008). Changes in gingival dimensions following connective tissue grafts for root coverage: comparison of two procedures. J Periodontol, 79(8), 1346-1354.
13. Henriques, P. S. G., Pelegri, A. A., Nogueira, A. A., & Borghi, M. M. (2010). Application of subepithelial connective tissue graft with or without enamel matrix derivative for root coverage: a split-mouth randomized study. J Oral Sci, 52(3), 463-471
14. Sculean, A., Cosgarea, R., Stahli, A., Katsaro, C., Arweiler, N. B., Brex, M., & Deppe, H. (2014). The modified coronally advanced tunnel combined with an enamel matrix derivative and subepithelial connective tissue graft for the treatment of a isolated mandibular Miller Class I and II gingival recession: a report of 16 cases. Quintessence Int, 45(10), 829-835
15. Cairo, F. (2017). Periodontal plastic surgery of gingival recessions at single and multiple teeth. Periodontol 2000, 75(1), 296-316
16. Imano, M. H., Chaves, L. H., Storrer, C. L. M., Amaral, C. F., Candido, B. F., & Deliberador, T. M. (2019). Use of Oxygen Gel as a Tissue Healing Optimizer in Donor and Recipient Areas in Free Gingival Graft Technique. IMPLANT NEWS PERIO; 1: 1.
17. Lafzi, A., Kadkhodazadeh, M., Mojahedi, S. M., Amid, R., Shidfar, S., & Baghani, M. T. (2019). The Clinical Evaluation of the Effect of Low-Level Laser Therapy on the Donor and Recipient Sites of the Free Gingival Graft: A Case Series. J Lasers Med Sci; 10(4):355-360. 10.15171/jlms.2019.58
18. Mattos, P. M., Papalexou, V., Tramontina, V. A., Kim, S. H., Lucyszyn, S. M., Bettega, P. V. C. et al. (2019). Evaluation of 2 techniques of epithelial removal in subepithelial connective tissue graft surgery: a comparative histological study. J Periodontal Implant Sci. 50(1):2-13. 10.5051/jpis.2020.50.1.2
19. Thoma, D. S., Gasser, T. J. W., Jung, R. E., & Hammerle, C. H. F. (2020) Randomized controlled clinical trial comparing implant sites augmented with a volume-stable collagen matrix or an autogenous connective tissue graft: 3-year data after insertion of reconstructions. J Clin Periodontol. 47(5):630-639. 10.1111/jcpe.13271

Quantitative analysis of IL 1 β level at Baseline, One week and One month

1. Group A(pg/ml)

S. No	T0	T1	T2
B1	82.4	96.4	58.2
B2	78.7	98.8	60.2
B3	80.6	97.2	58.4
B4	81.1	97.6	59.2

B5	79.2	97.2	58
Mean	80.4	97.4	58.8

2. Group B(pg/ml)

S. No	T0	T1	T2
C1	82.6	116.4	79.4
C2	79.5	114.6	78.3
C3	81.4	115.4	78.2
C4	82.4	115.1	78.8
C5	80.1	114.5	77.6
Mean	81.2	115.2	78.5

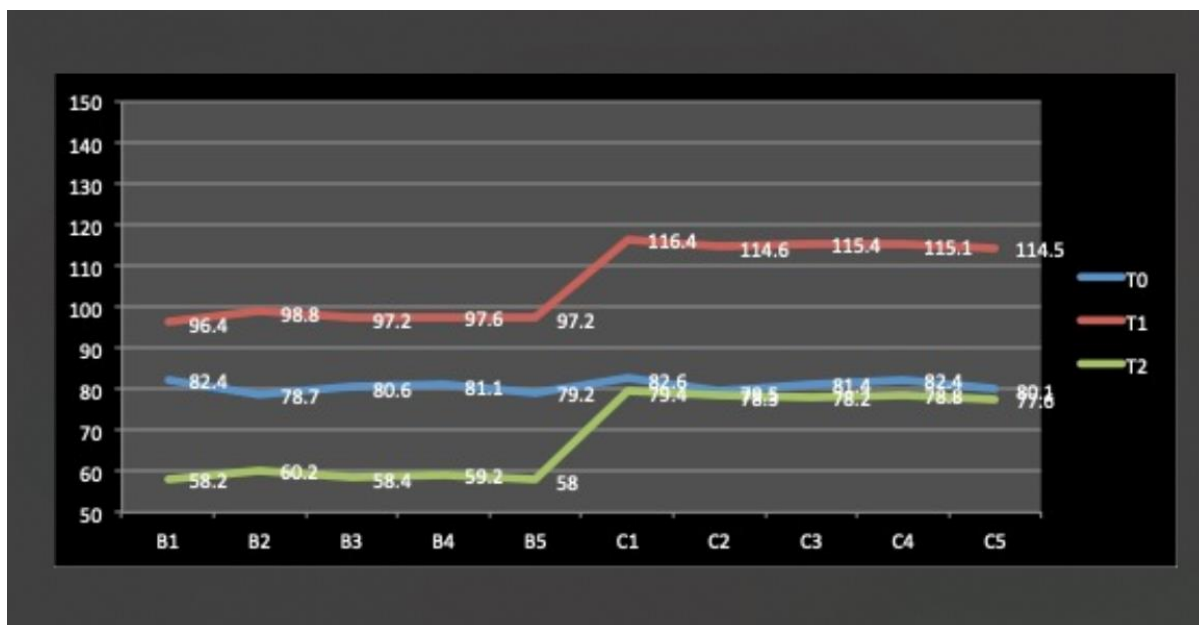


Figure 1