



**Molecular Basis of Tooth Movement**

<sup>1</sup>Ekta, PG Student, Department of Orthodontics and Dentofacial Orthopedics, K.D. Dental College, Mathura, Uttar Pradesh, India

<sup>2</sup>Atul Kumar Singh, Professor and HOD, Department of Orthodontics and Dentofacial Orthopedics, K.D. Dental College, Mathura, Uttar Pradesh, India

<sup>3</sup>Vipin Kumar Sharma, Reader, Department of Orthodontics and Dentofacial Orthopedics, K.D. Dental College, Mathura, Uttar Pradesh, India

<sup>4</sup>Ketki Dalvi, PG Student, Department of Orthodontics and Dentofacial Orthopedics, K.D. Dental College, Mathura, Uttar Pradesh, India

<sup>5</sup>Shivangi Mathur, PG Student, Department of Orthodontics and Dentofacial Orthopedics, K.D. Dental College, Mathura, Uttar Pradesh, India

**Corresponding Author:** Ekta, PG Student, Department of Orthodontics and Dentofacial Orthopedics, K.D. Dental College, Mathura, Uttar Pradesh, India

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**Abstract**

The connection between teeth and facial shape has been recognized since Biblical times, emphasizing the importance of dental alignment. Orthodontics, meaning "straightening teeth," addresses undesirable tooth positions through the application of controlled forces that induce remodeling of dental and paradental tissues, including the periodontal ligament (PDL), alveolar bone, and gingiva. Unlike physiological tooth drift, orthodontic tooth movement involves rapid changes in the PDL, creating distinct zones of compression and tension that affect vascularity and cellular activity.

Historically, orthodontics began with ancient recommendations for applying force to align teeth, evolving through significant contributions from figures like Pierre Fauchard and later hypotheses on bone resorption by Farrar and Wolff. Research in the 20th century focused on histological changes during tooth movement, revealing vital cellular activities in the PDL. Advancements in imaging and biochemical techniques have enhanced our understanding of the molecular and genetic responses of tissues to mechanical stress. This growing body of knowledge informs more biologically oriented and patient-friendly orthodontic practices. By clarifying the biological processes underpinning tooth

movement, clinicians can refine treatment approaches, ensuring effectiveness and safety, and paving the way for a future where orthodontics integrates biological principles for improved patient outcomes.

**Keywords:** Biological principles, Cellular activity, Molecular responses, Orthodontic tooth movement, Periodontal ligament (PDL) and Tooth drift.

### **Introduction**

Teeth play a crucial role in determining facial shape, a concept recognized since Biblical times. Orthodontics, meaning "straightening teeth," aims to correct undesirable tooth positions through the application of controlled forces. This movement induces remodeling in dental and paradental tissues, particularly the periodontal ligament (PDL), which responds dynamically to these forces. Unlike physiological tooth drift, orthodontic tooth movement creates specific zones of compression and tension that affect vascularity and cellular activity. A thorough understanding of these biological responses is essential for developing effective, safe, and patient-friendly orthodontic treatments, paving the way for more biologically oriented approaches in the field.

### **Cellular and Molecular Biology of Orthodontic Tooth Movement**

Orthodontic tooth movement (OTM) relies on applying appropriate forces to shift teeth through the alveolar bone, a process governed by mechanotransduction—the cellular response to mechanical stress. This interplay between biology and mechanics drives tissue remodeling in the dento-alveolar complex.

#### **Extracellular Matrix (ECM)**

The periodontal ligament (PDL), root cementum, and alveolar bone consist of cells and ECM, primarily made of collagen fibers and glycosaminoglycans (GAGs). Collagen type I predominates, providing tensile strength, while oxytalan fibers, lacking elastin, contribute to

elasticity. The ECM serves as a substrate for PDL cells, facilitating their migration and communication.

#### **Key Cell Types**

**Fibroblasts:** Central to matrix remodeling, they synthesize collagen and produce cytokines that regulate tissue responses and osteoclast activation.

**Osteoblasts:** Bone-forming cells derived from mesenchymal stem cells, they produce the organic bone matrix and play a role in mineralization.

**Osteocytes:** Inactive cells embedded in bone, they transmit signals that regulate osteoblast and osteoclast activity.

**Osteoclasts:** Multinucleated cells responsible for bone resorption, characterized by their ruffled borders that aid in mineral dissolution.

**Cementoblasts:** Derived from the dental follicle, these cells are involved in cementogenesis.

**Macrophages:** Key immune cells that remove debris and regulate tissue homeostasis.

#### **Phases of Orthodontic Tooth Movement**

Burstone proposed three phases of OTM: the initial phase (rapid movement), the lag phase (arrest due to tissue necrosis), and the post-lag phase (increased movement). Recent studies have refined this to four phases:

- 1. Initial Phase (24 hours to 2 days):** Rapid movement occurs as tooth displacement compresses the ECM, causing blood vessel occlusion and necrosis (hyalinization).
- 2. Arrest Phase (20-30 days):** Movement halts due to the presence of necrotic tissue, preventing osteoclast differentiation.
- 3. Acceleration Phase:** Begins with the removal of necrotic tissue, allowing tooth movement to resume.

**4. Linear Phase:** Tooth movement occurs at a constant rate, with resorption at the leading side and deposition at the trailing side.

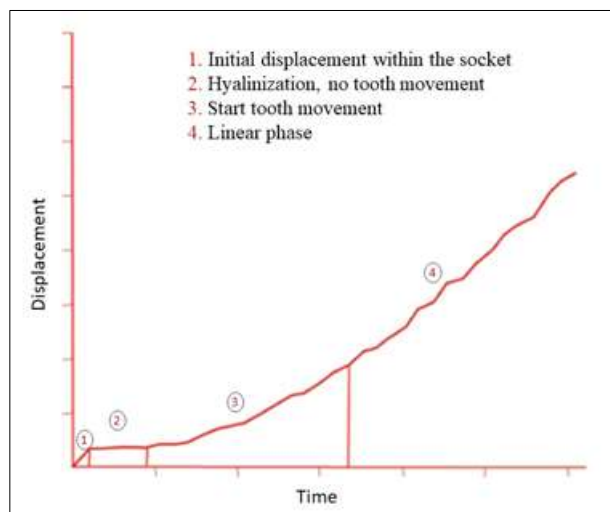


Figure 1: General time–displacement curve of OTM

#### Cellular Processes

During the initial phase, mechanical stress causes fluid redistribution, leading to hypoxia and cell death. Inflammatory responses initiate tissue repair. Osteoclasts migrate to necrotic areas for resorption. As the hyalinized tissue is cleared, new ECM is formed, primarily consisting of collagen type III. Mechanosensory osteocytes and fibroblasts respond to mechanical strain, promoting the release of growth factors and cytokines that facilitate the differentiation of precursors into osteoblasts and osteoclasts.

At the trailing side, the PDL widens, stimulating fibroblast proliferation and collagen synthesis. Growth factors like FGF-2 and VEGF enhance vascularization, ensuring optimal conditions for continued tooth movement.

#### Inflammatory Response in the Periodontal Ligament and Dental Pulp during Orthodontic Tooth Movement

Orthodontic tooth movement (OTM) involves the remodelling of the periodontal ligament (PDL) and alveolar bone, driven by mechanical forces that induce

an aseptic inflammatory response. This response is characterized by the release of various inflammatory mediators, including cytokines (e.g., IL-1, TNF- $\alpha$ ), neuropeptides, and damage-associated molecular patterns (DAMPs), which play crucial roles in the signalling pathways leading to tissue remodelling.

Cytokines such as IL-1 $\beta$  and TNF- $\alpha$  promote bone resorption and stimulate osteoclast activity through the RANK/RANKL/OPG system. Prostaglandins, particularly PGE<sub>2</sub>, also contribute to inflammation and bone remodeling. Furthermore, chemokines facilitate leukocyte migration to the site of inflammation, enhancing the local immune response.

Pain during OTM is a result of hyperalgesia triggered by inflammatory mediators, with symptoms beginning shortly after force application. Additionally, external apical root resorption (EARR) is a common complication linked to this inflammatory process, influenced by mechanical stress and cytokine signaling, particularly RANKL's role in osteoclastogenesis. Understanding these complex interactions is vital for optimizing orthodontic treatment and minimizing adverse effects.

#### Markers of Paradental Tissue Remodelling in Orthodontic Patients

**Oral Fluid Markers:** Analyzing gingival crevicular fluid (GCF) and saliva can help assess paradental tissue changes related to tooth movement.

**GCF vs. Saliva:** GCF, being less diluted, is more diagnostic for local inflammation than saliva, reflecting serum metabolite concentrations during orthodontic treatment.

**Inflammatory Response:** Orthodontic forces increase inflammation and capillary permeability, impacting fluid composition in GCF.

**Saliva Studies:** Short-term studies show increased salivary flow and cariogenic bacteria after orthodontic appliance placement, returning to baseline post-treatment.

**GCF Studies:** Specific markers identified in GCF include metabolic products, inflammatory mediators, and enzymes associated with bone remodeling.

**Collection Challenges:** GCF collection can vary based on inflammation, age, and smoking status; higher volumes noted in adolescents.

**Quantification Techniques:** Advanced methods like multiplex assays and Raman spectroscopy improve the analysis of GCF constituents.

**Future Directions:** Emerging technologies in “salivaomics” aim to enhance diagnostics for tooth movement, root resorption, and treatment outcomes, offering non-invasive and efficient sampling methods.

### **Effects of Drugs, Hormones, and Diet on Orthodontic Tooth Movement (OTM)**

**Tooth Movement Mechanisms:** OTM involves complex remodeling of paradental tissues influenced by inflammatory mediators, growth factors, and cytokines.

#### **Drug Effects on OTM:**

**Prostaglandins (PGs):** Enhance bone remodeling and tooth movement but can cause hyperalgesia. Clinical use limited due to short duration.

**NSAIDs:** Common pain relievers (e.g., ibuprofen) may reduce OTM if used chronically, but short-term use is less likely to affect movement.

**Antiresorptive Agents:** Bisphosphonates and denosumab significantly inhibit bone resorption, potentially slowing OTM. Caution advised during orthodontic treatment.

**Asthma Medications:** Certain medications may increase external root resorption during OTM; monitoring is essential.

**Corticosteroids:** Chronic use can lead to osteoporosis and may slow OTM; light forces recommended.

**Antihistamines:** May decrease OTM by inhibiting inflammation and bone resorption.

**Anticholinergic Drugs:** Can cause xerostomia, impacting oral hygiene and increasing caries risk.

**Psychiatric Medications:** Certain antidepressants may reduce the rate of tooth movement.

#### **Hormonal Influences on OTM:**

**Thyroid Hormones:** Can accelerate OTM; monitoring is advised for patients on therapy.

**Parathyroid Hormone:** Promotes bone density and does not negatively affect OTM during treatment.

**Estrogens:** May decrease OTM but often have minimal reported effects during orthodontic treatment.

#### **Nutritional Factors:**

**Vitamins:** Essential for metabolic processes; deficiencies (e.g., Vitamin C) can lead to periodontal issues affecting tooth mobility.

**Vitamin D:** Modulates calcium homeostasis and bone remodeling, positively impacting OTM.

**Minerals:** Copper and manganese are vital for collagen metabolism and bone health, though evidence on mineral supplementation effects on OTM is inconclusive.

### **Methods to Accelerate or Decelerate Orthodontic Tooth Movement**

#### **Biochemical Agents**

Cytokines, prostaglandins, RANKL, macrophage colony-stimulating factor, parathyroid hormone, vitamin D3, corticosteroids, and vascular endothelial growth factor facilitate tooth movement.

#### **Physical Agents**

Low-level lasers, direct electric currents, pulsed electromagnetic fields, vibrations, and surgical interventions are used to stimulate dental and paradental cells.

### Historical Attempts

- Early methods included heavy forces, which proved hazardous.
- Successful use of electric currents and prostaglandins to enhance tooth movement observed.

### Pharmacological Approaches

- Cytokine Delivery: Increases PDL cell differentiation and activation.
- Prostaglandins: Enhance tooth movement; injected locally, doubled movement rate.

### Hormones

- Parathyroid Hormone (PTH): Increases tooth movement; stimulates osteoclasts and osteoblasts.
- Vitamin D3: Increased osteoclasts and tooth movement in animal studies.
- Corticosteroids: Short-term effects negligible; long-term may induce bone resorption.

### Physical Stimuli

- **Electric Currents/Pulsed Electromagnetic Fields:** Some studies showed enhanced movement; low-quality evidence.
- **Vibratory Stimuli:** Increased tooth movement and osteoclast numbers observed in studies.
- **Photo Biomodulation:** Low-level laser therapy shown to enhance bone remodeling and tooth movement, but evidence remains weak.

### Surgical Approaches

- **Corticotomy:** Enhances tooth movement by reducing cortical bone resistance; mixed evidence on effectiveness.
- **Modifications:** Techniques like corticision and piezocision show promise but need further validation.

### Methods to Decelerate Tooth Movement

**Drugs:** Nonsteroidal anti-inflammatory drugs and bisphosphonates reduce osteoclast activity and tooth movement speed. Chemically modified tetracyclines and cetirizine also show potential in this regard.

### Conclusion

Orthodontics has evolved from simple mechanical methods to a sophisticated understanding of biological processes governing tooth movement. Research shows that tooth movement results from complex interactions between mechanical forces and biological responses in the periodontal ligament and alveolar bone. Key factors include inflammation, cytokines, and hormones that facilitate or inhibit osteoclast and osteoblast activity. Understanding these mechanisms allows for targeted strategies to accelerate or decelerate tooth movement, enhancing treatment efficiency. Personalized approaches based on individual biological profiles promise improved outcomes, addressing patient concerns about treatment duration and compliance in the growing adult orthodontic market.

### References

1. Krishnan, V. and Davidovitch, Z. (2009). Biological mechanisms of tooth movement.
2. Krishnan, V. and Davidovitch, Z. (2006) The cellular, molecular and tissue level reactions to orthodontic force. *AJO-DO* 129, e1–32.
3. Krishnan, V. and Davidovitch, Z. (2021). Biological mechanisms of tooth movement.
4. Schwarz, A. M. (1932). Tissue changes incidental to orthodontic tooth movement. *International Journal of Orthodontia, Oral Surgery and Radiography*, 18(4), 331–352. doi:10.1016/s0099-6963(32)80074-8.
5. Henneman, S., Von den Hoff, J. W., and Maltha, J. C. (2008). Mechanobiology of tooth movement.

- European Journal of Orthodontics 30, 299–306.  
doi.org/10.1093/ejo/ cjn020
6. Yamaguchi, M. (2009) RANK/RANKL/OPG during orthodontic tooth movement. *Orthodontics and Craniofacial Research* 12(2), 113–119. doi:10.1111/j.1601-6343.2009.01444.x.
  7. Grant, M., Wilson, J., Rock, P. and Chapple, I. (2013) Induction of cytokines, MMP9, TIMPs, RANKL and OPG during orthodontic tooth movement. *European Journal of Orthodontics* 35, 644–651.
  8. Rody, W. J., King, J. and Gu, G. (2001) Osteoclast recruitment to sites of compression in orthodontic tooth movement. *American Journal of Orthodontics and Dentofacial Orthopedics* 120, 477–489
  9. Meikle, M. C. (2006) The tissue, cellular, and molecular regulation of orthodontic tooth movement: 100 years after Carl Sandstedt. *European Journal of Orthodontics* 28, 221–240.
  10. Bumann, A., Carvalho, R. S., Schwarzer, C. L. and Yen, E. H. (1997) Collagen synthesis from human PDL cells following orthodontic tooth movement. *European Journal of Orthodontics* 9, 29–37.
  11. Krishnan, V. and Davidovitch, Z. (2006) The effect of drugs on orthodontic tooth movement. *Orthodontics and Craniofacial Research* 9, 163–171.
  12. Kyrkanides, S., Banion, K. O. and Subtelny, D. J. (2000) Non-steroidal anti-inflammatory drugs in orthodontic tooth movement – metalloproteinase activity and collagen synthesis by endothelial cells. *American Journal of Orthodontics and Dentofacial Orthopedics* 118, 203–209.
  13. Chumbley, A. B. and Tuncay, O. C. (1986) The effect of indomethacin (an aspirin like drug) on the rate of orthodontic tooth movement. *American Journal of Orthodontics and Dentofacial Orthopedics* 89, 312–314.
  14. Kehoe, M. J., Cohen, S. M., Zarrinnia, K. and Cowan, A. (1996) The effect of acetaminophen, ibuprofen and misoprotol on prostaglandin E2 synthesis and the degree and rate of orthodontic tooth movement. *The Angle Orthodontist* 66, 339–350.
  15. Kale, S., Kocadereli, I., Atilla, P. and Asan, E. (2004) Comparison of the effects of 1,25 dihydroxycholecalciferol and prostaglandin E2 on orthodontic tooth movement. *American Journal of Orthodontics and Dentofacial Orthopedics* 125, 607–614.