

Tumor Markers and Their Significance in Oral Dysplasia

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Abstract

Oral epithelial dysplasia (OED) denotes precancerous alterations in oral tissue, signalling potential malignancy. Not all cases of OED lead to oral squamous cell carcinoma (OSCC). The chances of progression depend on the severity of the dysplasia. Severe cases are more likely to progress, while mild cases can stay the same or

improve. Individual immune responses, genetic factors, and environmental exposures (e.g., tobacco and alcohol) impact the likelihood of progression, leading to diverse outcomes in each OED case. Tumor markers, whether produced by the tumor or as a response to its presence, play a crucial role in diagnosing cancer and evaluating the extent of the tumor. In OED, these markers are

critical for identifying high-risk patients, allowing early detection, timely intervention, and potentially improved treatment outcomes. However, the underutilization of these markers in predicting OED prognosis highlights the need for their more effective integration into clinical practice.

Incorporating these markers into clinical practice can enhance OED prognosis and management, ultimately benefiting individuals at risk for oral cancer.

Keywords: Oral squamous cell carcinoma, prognosis, treatment outcomes, epithelial-mesenchymal transition, cancer stem cells, Matrix Metallo Proteinases, Angiogenesis, oxidative stress, oncogenes.

Introduction

Oral epithelial dysplasia (OED) is a histopathological term used to identify a precancerous state within the oral epithelial tissue, now recognized as potentially malignant disorders. Its important to emphasize that not all cases of OED develop into oral squamous cell carcinoma (OSCC).^[1] OED is defined as an epithelial tissue in which the prevalence of OSCC is higher than in its healthy counterpart.^[2] The progression of OED into OSCC is a multifaceted process influenced by several factors. Not all instances of OED evolve into cancer due to various reasons including the degree of dysplasia, with severe dysplasia posing a higher risk of progression. In contrast, mild dysplasia may remain stable or regress. The individual's immune response is critical, as it can detect and eliminate precancerous cells. Genetic factors and environmental exposures, such as tobacco and alcohol, also impact the likelihood of progression.^[1] Regular monitoring and timely management of OED, along with lifestyle choices, contribute to preventing its transformation into cancer. Each case of OED is unique, and its variability further

contributes to the diverse outcomes observed in patients with this condition.

Tumor markers are substances that are produced either by the tumor itself or by the body in response to the presence of cancer or certain benign conditions that can aid in the diagnosis of cancer and the assessment of tumor burden.^[3, 4] Utilizing tumor markers in the prognosis of oral epithelial dysplasia is critically significant for the identification of high-risk patients. This, in turn, facilitates early detection, timely intervention, and the potential improvement of treatment outcomes and survival rates.

Nevertheless, the underutilization of these markers in predicting OED prognosis underscores the imperative for their more effective integration into clinical practice. Numerous studies have examined various immunohistochemical markers that are utilized to evaluate the malignant transformation of OED into OSCC. Several research studies have investigated a range of immunohistochemical markers that are employed to assess the progression of OED to OSCC.

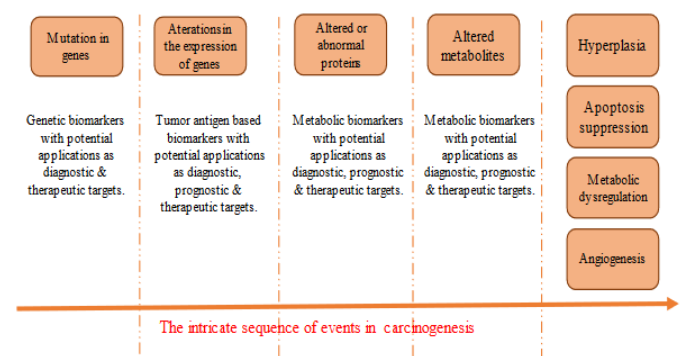


Figure- 1: The role of tumor markers in understanding and detecting carcinogenesis.

Ki-67

In 1983, Johannes Gerdes and colleagues introduced the monoclonal antibody Ki-67 (Antigen Kiel 67) as a marker for actively dividing cells. Ki-67 is effective in assessing cell proliferation, with advantages including resistance to external influences, minimal residual

staining, and specific nuclear expression during the cell cycle. Its valuable for evaluating the histologic grades of OED and OSCC, overexpressed in premalignant and malignant lesions.

Ki-67's positive cell fraction correlates with disease progression, with high-risk OED showing deeper proliferation into the basal layer compared to low-risk OED. [5, 6]

PCNA

Proliferating cell nuclear antigen (PCNA) is a pivotal protein crucial for DNA replication and repair processes. Acting as a cofactor for DNA polymerase delta, it plays a fundamental role in maintaining genomic stability. Additionally, PCNA serves as a reliable molecular marker for cell proliferation, with elevated expression during the G1 and S phases of the cell cycle. In OED, heightened PCNA levels indicate an increased cell proliferation index, creating a favourable environment for the accumulation of genetic alterations and potential cancer development. Consequently, monitoring PCNA expression emerges as a valuable prognostic tool for OED, signifying the potential for malignant transformation and emphasizing the necessity for sustained patient follow-up to identify any indications of possible malignancy. [7]

Cyclin D1

Oral carcinogenesis often involves disruptions in cell cycle control. Cyclin-dependent kinases (CDKs) and their inhibitors play a pivotal role in regulating the cell cycle. There are eleven cyclins, including D1-3 and E, which interact with CDK 4/6 and CDK 2, respectively, controlling the G1/S phase transition. Cyclin D1, located on chromosome 11q13, is a key regulator of the G1 phase. OED may exhibit molecular anomalies leading to cyclin D1 overexpression. [8-10] Cyclin D1 expression in oral dysplasia ranges from weak to strong in mild to

moderate cases, predominantly in the middle layer. Severe dysplasia exhibits strong expression across all layers, particularly increased in basal and parabasal layers. [11]

P27

The p27 gene is part of the kinase inhibitor protein family and inhibits CDKs. Initially, it was known for limiting G1 cyclin-CDK complexes, controlling cell proliferation. However, p27's role in carcinogenesis goes beyond proliferation control. Tumor p27 levels don't always align with the proliferative index, and subcellular localization, like cytoplasmic localization, can be a negative prognostic factor. Changes in p27 expression are tied to cell dynamics in OED, occurring before invasive stages of oral tumorigenesis. [12-16]

P21

Although p21 serves as a critical cyclin-dependent kinase inhibitor essential for cell cycle regulation, its precise role in OED remains elusive. Numerous investigations into p21 expression in both OED and OSCC present conflicting findings, with some studies indicating decreased levels in comparison to normal oral mucosa, while others report increased expression. This variability suggests a multifaceted and context-dependent role for p21 in OED and OSCC. One plausible explanation for reduced p21 expression could be linked to the inactivation of the p53 tumor suppressor pathway. As a downstream target of p53, p21 expression is induced by p53 in response to DNA damage or stress signals, potentially resulting in decreased expression if p53 function is compromised in OED and OSCC. Conversely, elevated p21 expression in these conditions may be attributed to the activation of alternative pathways governing p21 expression. Examples include the Akt/mTOR pathway, which some studies propose as a contributor to increased p21 expression in OSCC.

Additionally, cell cycle phase accumulation, particularly in the G1 phase, has been suggested as a factor associated with heightened p21 expression in OED and OSCC. The intricate dynamics of p21 in these conditions underscore the need for further research to unravel the underlying mechanisms and assess the potential utility of p21 as a diagnostic or prognostic marker.^[17]

MDM2

MDM2 (Murine double minute 2), an oncogene, acts as a negative regulator of the p53 tumor suppressor pathway, frequently compromised in OED and OSCC. MDM2's role involves promoting p53 degradation through ubiquitination and subsequent proteasomal breakdown, potentially contributing to p53 inactivation in OED and OSCC. Studies on MDM2 expression in these conditions present conflicting findings, with some reporting increased expression compared to normal oral mucosa and others finding no significant difference. The relationship between MDM2 expression and clinical outcomes remains unclear. Recent research suggests a potential therapeutic role for MDM2 in cancer treatment. Anti-MDM2 antisense oligonucleotides have shown promising in vitro and in vivo activities against various human cancer models, inducing p21 regardless of p53 status. However, the complex role of MDM2 in OED and OSCC requires further investigation to unravel dysregulation mechanisms and assess its potential as a diagnostic, prognostic marker, or therapeutic target.^[17]

p53

P53, also known as tumor protein 53, plays a crucial role in maintaining cellular balance, and its mutations are often linked to factors such as alcohol and tobacco use. Various studies have established a positive connection between p53 expression and tumor progression. However, it's worth noting that overexpression of this protein can be influenced by the location of lesions,

being more common in areas with high sun exposure. Additionally, the presence of p53 protein is absent in squamous cell carcinomas but is observed in patients who have undergone immunosuppressive treatments, like organ transplant recipients. This suggests that the immune system also regulates the expression of this protein. In premalignant oral lesions, increased p53 expression typically correlates with the degree of epithelial dysplasia, indicating an elevated risk of malignant transformation. Many research teams have concluded that in the case of premalignant oral lesions, the level of p53 expression directly correlates with the risk of developing squamous cell carcinoma, even after long-term monitoring of these lesions' progression.^[18-20]

Immunohistochemical patterns correlated with P53 mutations and high SCC risk, emphasizing the need for improved diagnosis beyond WHO classification.^[21]

Her2

The human epidermal growth factor receptor-2/neu (ErbB) protein, a member of the tyrosine kinase receptor family, plays a crucial role in cellular growth and differentiation. Dysregulation of HER-2/neu is associated with uncontrolled cell cycle, resistance to apoptosis, invasiveness, chemo-resistance, and angiogenesis. Overexpression of HER-2/neu has been linked to poor clinical outcomes in various cancers, including breast, ovarian, osteosarcoma, endometrial, salivary, and gastric carcinomas.

Studies on HER-2/neu in OSCC show variable and discordant results, with incidence ranging from 0 to 40%. Targeted therapies against HER-2/neu have shown efficacy in breast, gastric, and lung cancers.^[22] Evaluating HER-2/neu expression in OPMDs and OSCC will help identify a potential biomarker for specific immunotherapy against OSCC, suggesting a valuable avenue for further clinical investigation.

Rb

The retinoblastoma (Rb) tumor suppressor gene regulates cell cycle progression and differentiation through its protein product, Retinoblastoma tumor suppressor protein (pRb). pRb acts as a regulator at the G1-S restriction point, inhibiting growth in mid G1-S phase. Mutations lead to functional pRb inactivation and failure of tumor suppression. In oral cancer, there is evidence of dysfunctional Rb protein pathways. In oral cancer, abnormal Rb protein types or reduced Rb production may lead to low availability of functional pRb. Rb plays a crucial role in terminal differentiation, influencing cell cycle withdrawal and gene modulation for a differentiated phenotype.^[23] pRb is considered a tumor suppressor protein rather than a specific molecular marker. Tumor suppressor proteins, like Rb, play a crucial role in regulating cell cycle progression and preventing the development of tumors. This protein is not typically used as a diagnostic marker in the same way as specific molecular markers but is studied for its involvement in cancer development and progression.

Stathmin

Stathmin plays a crucial role in regulating the microtubule cytoskeleton by influencing microtubule dynamics. It facilitates the depolymerization of microtubules and inhibits the polymerization of tubulin heterodimers. This depolymerization process involves sequestering free tubulin dimers or directly inducing microtubule catastrophe. Numerous studies on stathmin indicate its frequent overexpression in various human cancers, highlighting its close correlation with cancer cell differentiation, TNM classification, and lymph node metastases. The observed overexpression of stathmin is linked to sustained cancer cell proliferation, underscoring its significance in tumorigenesis and tumor development.

Stathmin serves as a valuable oncobiological marker and a potential molecular target for cancer therapy.^[24]

S100

The S100 protein family, with its diverse distribution in the cytoplasm and cell organelles, plays a multifaceted role in cellular functions, largely attributed to its calcium-binding properties. In the context of oral epithelial dysplasia, S100's functions include regulation of cell proliferation, cell division, Ca^{2+} homeostasis, structural organization of membranes, dynamics of cytoskeleton constituents, cell growth, motility, survival, enzyme activation, protection from oxidative damage, and involvement in protein phosphorylation and secretion. While intracellularly, S100 controls the cell cycle, extracellularly, it acts as a cytokine binding to receptors like RAGE and TLRs. In cancer, dysregulation of S100 proteins is linked to differentiation, with varied expressions observed in different members of the S100 family. Notably, S100A14 has been implicated in modulating the expression of MMP1 and MMP9 in invasive tumor cells of OSCC, providing insights into its potential role in the prognosis of OED.^[25]

P16

P16, or MTS1 (multiple tumor suppressor 1), is a tumor suppressor that interacts with p21, CDK, and PCNA. It regulates cell proliferation, differentiation, and apoptosis independently of p53. In OSCC cases originating from precursor lesions, about 24% show mutations in the gene responsible for p16 production.^[26,27] Post-transplant immunosuppressed patients exhibit p16 overexpression in premalignant and malignant lesions. Methylated p16 might serve as a marker for malignancy and is found in premalignant lesions and neighbouring epithelium. Moreover, p16 overexpression may be associated with an infectious cause related to human papillomavirus (HPV)^[28]

EGFR

Cytogenetic analysis in head and neck tumors reveals oncogene activation, tumor suppressor gene inactivation, and growth factor expression, including EGF, TGF- α , TGF- β , and FGF, along with EGFR receptors. EGFR, overexpressed in oral cancers, has become a promising target for monoclonal antibody therapy. Its high expression is linked to tumor size, metastasis, and survival. Exploring mutations in the EGFR 2 gene can provide insights into its overexpression in oral cancer.

Tumors with high EGFR expression respond better to chemotherapy, possibly due to increased proliferative activity. Early overexpression of TGF- α in oral carcinogenesis, along with EGFR, may indicate more aggressive tumors. [29, 30]

Survivin

Survivin, a member of the inhibitor of apoptosis (IAP) protein family, holds significance in cancer due to its role in regulating mitosis and apoptosis. Highly expressed in various malignancies, including OSCC, survivin is associated with aggressive disease and poor clinical outcomes. Studies on survivin gene polymorphisms indicate their potential utility in disease prognosis and diagnosis, influencing tumor aggressiveness and patient survival. Survivin's differential expression in cancer cells, coupled with its involvement in key cellular pathways, positions it as a promising target for therapeutic interventions. In the context of OED, survivin's intricate role and potential as a diagnostic or prognostic marker necessitate further investigation. [31]

HIF-1 α

In oral carcinogenesis, HIF-1 α plays a pivotal role in orchestrating various pathways that contribute to a more aggressive tumor phenotype, characterized by metastases and the promotion of epithelial-mesenchymal transition (EMT). HIF-1 α is a central transcription factor involved

in the metabolic adaptation of cancer cells, particularly in the shift from oxidative phosphorylation to glycolysis. The stabilization of HIF-1 α , both under hypoxic and normoxic conditions, is associated with alterations in glucose and glutamine metabolism, leading to increased energy consumption for enhanced tumor cell proliferation. Additionally, HIF-1 α upregulates key proteins such as carbonic anhydrase IX (CA IX), hexokinase 2 (HK2), and various proton exchangers, contributing to the acidification of the tumor microenvironment. The lowered extracellular pH serves as a driving force in the early stages of EMT, further promoting cancer aggressiveness. Numerous studies have highlighted the prognostic significance of HIF-1 α expression in OSCC, suggesting its potential utility as an independent prognostic marker for patient outcomes. [32]

GLUT1 & HK2

Glucose metabolism is essential for cellular energy production, and its dysregulation is associated with tumorigenesis. Glucose transporters (GLUTs) facilitate glucose entry into cells, and hexokinases (HKs), particularly HK-II, are key enzymes in glycolysis. In OED progression, the overexpression of GLUT-1 and HK-II is observed, contributing to the Warburg effect, a phenomenon where cancer cells favor glycolysis for energy production. This heightened glycolytic activity is crucial for the increased energy demands of rapidly proliferating malignant tumors. Aberrant expression of GLUT-1 and HKII is linked to invasion and metastasis in head and neck cancers, including OED. Furthermore, their abnormal expression is associated with the hypoxic tumor microenvironment. Inhibition of GLUT-1 and HK-II has shown promise in improving the treatment efficacy of malignant tumors, suggesting a potential

novel therapeutic strategy for head and neck cancers, including OED. [33]

β-catenin

In oral epithelial dysplasia, β-catenin's dynamic subcellular localization serves as a pivotal factor in understanding the progression towards oral cancer. There is an intriguing pattern where dysplastic oral keratinocytes exhibit high nuclear accumulation of β-catenin, contrary to the cytoplasmic predominance observed in oral carcinoma. Unlike some malignancies, mutations in Wnt/β-catenin pathway components are not prevalent in oral carcinogenesis. Instead, the overproduction of Wnt ligands, notably Wnt3a, emerges as a potential explanation for aberrant pathway activation. This heightened Wnt/β-catenin activity, facilitated by increased Wnt ligand release, may contribute to the proliferation of dysplastic cells. Moreover, epigenetic silencing of Wnt inhibitors and the potential involvement of pathways like PI3K-Akt and EGFR warrant further exploration. Understanding the intricate role of β-catenin in dysplasia could offer insights into prognosis and therapeutic interventions for oral precancerous lesions. [34]

PTEN

PTEN (Phosphatase and tensin homologue deleted on chromosome 10) is a tumor suppressor gene rather than a direct marker. As a tumor suppressor gene, PTEN encodes a protein that plays a critical role in regulating cell growth, proliferation, and survival by inhibiting the PI3K/AKT signaling pathway. PTEN acts as a phosphatase, dephosphorylating phosphatidylinositol triphosphate (PIP3), which leads to the activation of AKT (Protein kinase B). The loss or inactivation of PTEN function is often associated with various cancers, including OSCC, and is implicated in tumorigenesis. In this context, PTEN serves as a marker for dysregulation

of the PI3K/AKT pathway and a potential indicator of malignant transformation of OED. [35]

Matrix Metallo Proteinases

Matrix metalloproteinases (MMPs) degrade the extracellular matrix during tumor progression. High levels of MMP-1, MMP-2, and MMP-9 in severe dysplasia are linked to poor prognosis. Notably, MMP-2, normally absent in healthy mucosa, increases with dysplasia. MMP-9 shows promise as a prognostic marker in oral cancer, with elevated levels in lesions with malignant potential. However, its role as a reliable biomarker requires additional research. [36]

Vimentin

Vimentin, traditionally a mesenchymal structural protein, now regulates cellular functions, impacting migration, signalling, adhesion, and tumor metastasis. Overexpression, notably in OSCC, correlates with unfavourable outcomes. Molecular markers' localization at invasive fronts holds significant prognostic value. Abnormal vimentin expression strongly links to tumor invasion and metastasis, with limited exploration in precancerous lesions, except for its confirmation in hyperkeratotic oral mucosal lesions. [37-40]

E-cadherin

E-cadherin, a 120-kDa glycoprotein, facilitates cell adhesion and inhibits proliferation in epithelial tissues. Decreased E-cadherin expression is linked to aggressiveness and poor outcomes in various carcinomas. In OED, reduced E-cadherin predicts higher disease risk, invasion, and a worse prognosis, making it a vital prognostic biomarker. [41-43] In a study by Nagireddi Puneeta et al. in oral dysplasia, E-cadherin expression decreases progressively from mild (90%) to severe (70%). Notably, there is a significant

shift from membranous to cytoplasmic localization with increasing dysplasia severity. [44]

Bcl-2

Cancer arises when mutations disrupt the regulation of apoptosis and cell survival mechanisms. These processes are controlled by various gene sets, with the Bcl-2 family being prominent. This family includes both pro- and anti-apoptotic members that share sequence homology in their Bcl-2 homology domains. Bcl-2, initially identified in human follicular non-Hodgkin's B-cell lymphomas, is a proto-oncogene. The Bcl-2 protein, weighing 26 kDa, is found in the nuclear envelope, endoplasmic reticulum, and the outer mitochondrial membrane, encoded by chromosome 18q21. Bcl-2's role in prolonging cell survival can lead to the accumulation of additional mutations in other tumor suppressor genes and oncogenes, contributing to clonal progression. [45-48]

c-myc

The c-myc gene plays a pivotal role in promoting transcription and cellular proliferation through cyclin-dependent kinase initiation. In HNSCC, C-Myc influences EMT by altering adhesion proteins and inducing MMPs, notably MMP-9. The progression of HNSCC is further influenced by the collaboration of C-Myc with genes like cyclin D1 and hypoxia-inducible factor (HIF)-related genes such as VEGF. The BCL2 gene, known for its role in inhibiting cell death through the Bcl-2 protein, parallels c-myc in cancer development. [49]

Fascin

Fascin, an up-regulated actin cross-linking protein in various carcinomas, crucially influences the prognosis of OED. Primarily expressed in motile cells, it orchestrates the organization of cellular protrusions, contributing to cell migration and the progression of carcinomas. In oral cancer, fascin's tight bundling of F-actin and early up-

regulation play a pivotal role in the transition from normal epithelium to invasive OSCC, underscoring its significance as a prognostic marker for OED. [50]

COX-2/PGE2

COX-2/PGE2 plays a crucial role in oral epithelial dysplasia. In this context, inflammation induced by COX-2/PGE2 stimulates the division of initiated tumour cells, inhibits cell death, and promotes angiogenesis, allowing the tumour to grow. This process involves the production of cytokines, ROS, and activation of transcription factors like NF-κB and STAT3. Ultimately, COX-2/PGE2 contributes to the progression of OED through multiple mechanisms, including oxidative stress, immune suppression, and angiogenesis. [51]

VEGF

Solid tumors, constrained to 1–2 mm³ without ample blood supply, necessitate angiogenesis for growth, invasion, and metastasis. [52] In the oral cavity, 10–20% of dysplastic lesions progress to carcinomas. [53] Studies reveal increased vascularity from normal oral mucosa to invasive carcinoma during dysplasia progression. Significantly, a robust association between tumor angiogenesis and the progression of OSCCs is noted. Vascular endothelial growth factor (VEGF) assumes a crucial role in fostering angiogenesis across various tumor types, with numerous studies extensively exploring its involvement in the context of oral dysplasia and cancers. [54-57]

CK19

CK19 expression varies in oral conditions like normal, hyperplasia, dysplasia, and OSCC across studies. Unlike some reports, normal oral mucosa shows CK19 in both keratinized and nonkeratinized epithelium. Focal epithelial hyperplasias display inconsistent CK19 expression in association with inflammation. Dysplasia

challenges the expected progressive increase, with a decline in mild and moderate cases and a significant rise in severe dysplasia. OSCC's CK19 expression is inconsistent in indifferent studies. These findings highlight CK19's complex role in oral conditions. Further exploration of CK19 as a prognostic marker in oral epithelial dysplasia is warranted, considering the variations observed.^[58-61]

CD44

CD44, implicated in cancer cell migration, is cleaved by MT1-MMP. TGF- β induces MT1-MMP. CD44 is a CSC marker in head and neck SCC, contributing to drug resistance and EMT. CD44's pleiotropic role includes interaction with hyaluronate, fibronectin, and collagen, crucial for cell-cell/matrix interactions, migration, and tumor progression. The EMT process is vital for various physiological processes and correlates with the invasive behavior of cancers like head and neck SCC.^[62]

SOX2

SOX2, implicated in OSCC, exhibits varied expression in OPMDs like leukoplakia. Its overexpression, observed in OPMDs, suggests a potential role in OSCC pathogenesis. However, conflicting findings associate SOX2 with both poor and improved prognosis in OSCC, necessitating further exploration for accurate prognosis in oral epithelial dysplasia.^[63]

CD133

CD133, also known as Prominin-1, has emerged as a pivotal marker with significant implications in the prognosis of OED, OSCC and other malignancies. CD133, characterized by an N-terminal extracellular domain, five transmembrane domains, and an intracellular cytoplasmic tail with functional tyrosine kinase sites serves as a surface marker for Cancer Stem Cells (CSCs). Originally identified as a hematopoietic stem cell marker (Ravindran and Devaraj, 2012), CD133

has evolved into a key CSC marker for solid tumors, encompassing breast, gastric, pulmonary, hepatic, prostate, pancreas, and thyroid cancers. In the context of OED, CD133's significance lies in its ability to identify and characterize CSCs, which play a crucial role in tumor initiation and progression. Its presence as a marker suggests a potential association with the aggressiveness and prognosis of OSCCs. As CD133 is linked to distinct cytoplasmic interactions and possesses tyrosine kinase sites, it likely participates in intricate signalling pathways that influence the behavior of cancer cells.^[64]

OCT 4

The OCT4 gene (octamer binding transcription 4), essential for regulating early-stage growth and development, undergoes decreased expression following cell differentiation and maturation. In OSCC, positive OCT4 immunohistochemical expression is linked to significantly inferior overall survival rates. Particularly, the prognosis becomes notably unfavourable in cases of triple-positive OSCC (OCT4/NANOG/CD133), underscoring the adverse impact of these combined markers on outcomes.^[65]

NANOG

NANOG, a key regulator in the development and progression of OSCC, emerges as a significant factor in the prognosis of OED. NANOG expression is notably absent in normal tissue but is significantly upregulated in OSCC, especially in cases with nodal metastases, suggesting its potential as a prognostic biomarker. Immunohistochemical analysis of NANOG usually reveals positive staining in a substantial proportion of OSCC cases, displaying a cytoplasmic pattern distinct from the nuclear pattern observed in germ-cell tumors. While the significance of these different staining patterns

remains unclear, their presence in dysplastic epithelium adjacent to OSCC further emphasizes the role of NANOG in tumor development. Despite its crucial role in OSCC, the regulation of NANOG expression is not fully understood. The positive correlation between NANOG and mRNA levels of selected protein regulators (AGR2, KLF4, NOTCH1, OCT4, and SOX2) in tumor tissue samples underscores their influence on NANOG expression in OSCC. [66]

BAX

Bax (Bcl2 associated x protein), identified as a pro-apoptotic member of the Bcl-2 protein family, plays a critical role in governing programmed cell death or apoptosis. Specifically implicated in the intrinsic pathway of apoptosis, activated by cellular stresses such as DNA damage and oxidative stress, Bax undergoes a structural alteration and relocates to the mitochondria. There, it facilitates the release of cytochrome c and other pro-apoptotic factors, initiating caspase activation and eventual cell demise.

In the context of OED, a decline in Bax expression signifies potential dysregulation in apoptosis mechanisms, hindering the elimination of genetically damaged cells and escalating the risk of malignant transformation. Several studies propose that this reduction in Bax expression is pivotal in the onset and advancement of oral cancer, underscoring its significance in understanding the underlying mechanisms influencing the transformation of oral epithelial cells. [7]

Laminin and fibronectin

In OSCC, variations in laminin and fibronectin isoforms play a crucial role in cellular processes such as adhesion and motility. The basement membrane (BM) composition changes during cellular maturation, with fetal oral squamous epithelium containing laminin chains $\alpha 2$, $\alpha 3$, $\alpha 5$, $\beta 1$, $\beta 2$, $\beta 3$, $\gamma 1$, and $\gamma 2$, while adult

normal oral squamous epithelium comprises $\alpha 3$, $\alpha 5$, $\beta 1$, $\beta 3$, $\gamma 1$, and $\gamma 2$. Adult hyperproliferative, dysplastic, and carcinomatous lesions show re-expression of laminin $\alpha 2$ and $\beta 2$ chains. OED and OSCC exhibit BM breaks, correlating with malignancy grade. In the invasion front, laminin-5 chains $\alpha 3$ and $\gamma 2$ are found outside the BM, suggesting laminin-5 as an immunohistochemical marker for invasion and indicating a guiding role in OSCC invasion. Oncofetal fibronectins, specifically ED-B fibronectin, are expressed in the stromal compartment, particularly in areas of desmoplasia where stromal myofibroblasts contribute to the fetal extracellular matrix milieu. Overall, OSCC demonstrates a fetal extracellular matrix conversion involving both tumor cells and recruited stromal myofibroblasts. [67]

Conclusion

OED serves as a critical precursor to OSCC. The significance lies in tumor markers, which are powerful tools for identifying high-risk OED patients and facilitating timely intervention. However, the underutilization of these markers underscores a missed opportunity for predicting OED prognosis.

Their effective integration into clinical practice is essential. Embracing tumor markers can transform OED management, offering a lifeline to individuals at risk of developing oral cancer, emphasizing the vital role they play in early detection and improved outcomes.

Table 1

Markers based on their roles in evaluating the progression of OED to OSCC.		
Marker	Low-Risk Patients	High-Risk Patients
Proliferation Markers		
Ki-67	Decreased	Increased
PCNA	Decreased	Increased
Cell Cycle Regulation		
Cyclin D1	Decreased	Increased
P27	Increased	Decreased
P21	Increased/ Decreased	Increased/ Decreased
MDM2	Decreased	Increased
P53	Decreased	Increased
HER2	Decreased	Increased
Rb	Decreased	Increased
Stathmin	Decreased	Increased
S100	Decreased	Increased

Genetic and Molecular Markers		
p16	Increased	Decreased
EGFR	Increased	Decreased
Survivin	Increased	Decreased
HIF-1 α	Increased	Decreased
GLUT-1	Increased	Decreased
HK2	Increased	Decreased
β -catenin	Increased	Decreased
PTEN	Increased	Decreased
Matrix Metalloproteinases Enzymes		
MMP 1	Decreased	Increased
MMP 2	Decreased	Increased
MMP 9	Decreased	Increased
Epithelial-to-Mesenchymal Transition (EMT) Markers		
Vimentin	Increased	Decreased
E-cadherin	Increased	Decreased
c-myc	Increased	Decreased
Fascin	Increased	Decreased
Cell Adhesion and Migration		
Bcl2	Decreased	Increased
Angiogenesis and Inflammation Markers		
COX-2/PGE2	Increased	Decreased
VEGF	Increased	Decreased
VEGF	Increased	Decreased
Differentiation and Tumor Initiation		
CK19	Decreased	Increased
Cancer Stem Cell Markers		
CD44	Increased	Decreased
SOX2	Increased	Decreased
CD133 (Prominin 1)	Increased	Decreased
OCT4	Increased	Decreased
NANOG	Increased	Decreased
Pro-apoptotic Markers		
P53	Increased	Decreased
BAX	Increased	Decreased
Matrix Proteins		
Laminin	Decreased	Increased
Fibronectin	Decreased	Increased
* These markers collectively aid in evaluating the prognosis and risk of malignant transformation in OED, with each category playing a distinct role in the assessment.		

References

1. Nevanpää, T.T., Terävä, A.E., Laine, H.K. et al. Malignant transformation of oral epithelial dysplasia in Southwest Finland. Sci Rep 12, 8261 (2022). <https://doi.org/10.1038/s41598-022-12441-9>
2. El-Naggar, A.K.C.J. et al. WHO Classification of Head and Neck Tumours, 4th edn. (International Agency for Research on Cancer, Lyon, 2017).
3. Sanjay BR, Madhavi BR, Shyam NDVN. Tumour markers in oral neoplasia. IJDA 2010;2:103-14.
4. Sultana NS, Sham E, Kaul R, Shastri S, Bhat S. Tumor markers: A short overview. Int J Oral Maxillofac Pathol 2013;4:7-15
5. Ross W, Hall PA. Ki67: From antibody to molecule to understanding? Clin Mol Pathol 1995;48:M113-7
6. Birajdar SS, Radhika MB, Paremala K, Sudhakara M, Soumya M, Gadivan M. Expression of Ki-67 in normal oral epithelium, leukoplakic oral epithelium and oral squamous cell carcinoma. J Oral Maxillofac Pathol 2014;18:169-76.
7. Fernando A. C. G. de Sousa, Thaís C. Paradella, Yasmin R. Carvalho, and Luiz E. B. Rosa. Immunohistochemical expression of PCNA, p53, bax, and bcl-2 in oral lichen planus and epithelial dysplasia. Journal of Oral Science, 2009, 51(1), 117-121. DOI: 10.2334/josn.51.117. Sherr CJ. G1 phase progression: Cycling on cue. Cell 1994;79:551-5
8. Poon RY. "Cell cycle control". In: Encyclopedia of Cancer. Bertino JR, editor. San Diego: Academic Press; 1997. p. 246-55.
9. Huang S, Chen CS, Ingber DE. Control of Cyclin D1, p27Kip1, and cell cycle progression in human capillary endothelial cells by cell shape and cytoskeletal tension. Mol Biol Cell 1998;9:3179-93
10. Ramasubramanian A, Ramani P, Sherlin HJ, Premkumar P, Natesan A, Thiruvengadam C. Immunohistochemical evaluation of oral epithelial dysplasia using cyclin-D1, p27 and p63 expressions as predictors of malignant transformation. J Nat Sci Biol Med 2013;4:349-58.
11. Polyak K, Kato JY, Solomon MJ, Sherr CJ, Massague J, Roberts JM, et al. p27Kip1, a Cyclin-Cdk inhibitor, links transforming growth factor beta and contact inhibition to cell cycle arrest. Genes Dev 1994;8:9-22.

12. Singh SP, Lipman J, Goldman H, Ellis FH Jr, Aizenman L, Cangi MG, et al. Loss or altered subcellular localization of p27 in Barrett's associated adenocarcinoma. *Cancer Res* 1998;58:1730-5.
13. Tan P, Cady B, Wanner M, Worland P, Cukor B, Magi-Galluzzi C, Lavin P, Draetta G, Pagano M, Loda M. The cell cycle inhibitor p27 is an independent prognostic marker in small (T1a, b) invasive breast carcinomas. *Cancer research*. 1997 Apr 1;57(7):1259-63.
14. Jordan R, Bradley G, Slingerland JM. Reduced levels of the cell-cycle inhibitor p27Kip1 in epithelial dysplasia and carcinoma of the oral cavity. *Am J Pathol* 1998;152:585-90
15. Thambiah LJ, Bindushree RV, Anjum A, Pugazhendhi SK, Babbu L, Nair RP. Evaluating the expression of p16 and p27 in oral epithelial dysplasias and oral squamous cell carcinoma: A diagnostic marker for carcinogenesis. *J Oral Maxillofac Pathol*. 2018 Jan-Apr;22(1):59-64. doi:10.4103/jomfp.JOMFP_92_17. PMID: 29731558; PMCID: PMC5917543.
16. Perez-Ordóñez B, Beauchemin M, Jordan RC. Molecular biology of squamous cell carcinoma of the head and neck. *J Clin Pathol*. 2006;59:445–453. doi: 10.1136/jcp.2003.007641.
17. Zhang Z, Wang H, Li M, Agrawal S, Chen X, Zhang R. MDM2 Is a Negative Regulator of p21WAF1/CIP1, Independent of p53. *Journal of Biological Chemistry*. 2004 Apr;279(16):16000-16006. doi: 10.1074/jbc.M312264200.
18. Chang KW, Lin SC, Kwan PC, Wong YK. Association of aberrant p53 and p21 (WAF1) immunoreactivity with the outcome of oral verrucous leukoplakia in Taiwan. *J Oral Pathol Med*. 2000;29:56–62. doi: 10.1034/j.1600-0714.2000.290202.x.
19. Calenic B, Greabu M, Caruntu C, Nicolescu MI, Moraru L, Surdu-Bob CC, Badulescu M, Anghel A, Logofatu C, Boda D. Oral keratinocyte stem cells behavior on diamond like carbon films. *Rom Biotechnol Lett*. 2016;21:11914–11922.
20. Sawada, K., Momose, S., Kawano, R. et al. Immunohistochemical staining patterns of p53 predict the mutational status of TP53 in oral epithelial dysplasia. *Mod Pathol* 35, 177–185 (2022). <https://doi.org/10.1038/s41379-021-00893-9>
21. Lu S, Tiekso J, Hietanen S, Syrjänen K, Havu VK and Syrjänen S: Expression of cell-cycle proteins p53, p21 (WAF-1), PCNA and Ki-67 in Benign, premalignant and malignant skin lesions with implicated HPV involvement. *Acta Derm Venereol* 79: 268-273, 1999
22. Mirza S, Hadi N, Pervaiz S, Zeb Khan S, Mokeem SA, Abduljabbar T, Al-Hamoudi N, Vohra F. "Expression of HER-2/neu in Oral Squamous Cell Carcinoma." **Asian Pac J Cancer Prev.** 2020 May 1;21(5):1465-1470
23. Thomas S, Balan A, Balaram P. The expression of retinoblastoma tumor suppressor protein in oral cancers and precancers: A clinicopathological study. *Dent Res J (Isfahan)*. 2015 Jul-Aug;12(4):307-14. doi: 10.4103/1735-3327.161427. PMID: 26288619; PMCID: PMC4533187.
24. Vadla P, Yeluri S, Deepthi G, Guttikonda VR, Taneeru S, Naramala S. Stathmin! An immunohistochemical analysis of the novel marker in Oral Squamous Cell Carcinoma and Oral Leukoplakia. *Asian Pacific Journal of Cancer Prevention: APJCP*. 2020 Nov;21(11):3317.

25. Kuberappa PH, Bagalad BS, Ananthaneni A, Kiresur MA, Srinivas GV. Certainty of S100 from Physiology to Pathology. J Clin Diagn Res. 2016 Jun;10(6):ZE10-5. doi:10.7860/JCDR/2016/17949.8022. Epub 2016 Jun 1. PMID: 27504432; PMCID: PMC4963792.
26. Presland RB and Jurevic RJ: Making sense of the epithelial barrier: What molecular biology and genetics tell us about the functions of oral mucosal and epidermal tissues. J Dent Educ 66: 564-574, 2002
27. Condurache Hritcu OM, Botez AE, Olinici DT, Onofrei P, Stoica L, Grecu VB, Toader PM, Gheucă-Solovăstru L, Cotrutz EC. Molecular markers associated with potentially malignant oral lesions (Review). Exp Ther Med. 2021 Aug;22(2):834. doi: 10.3892/etm.2021.10266. Epub 2021 Jun 4. PMID:34149880; PMCID: PMC8200803.
28. Rajeswari MR, Saraswathi TR. Expression of epithelial growth factor receptor in oral epithelial dysplastic lesions. J Oral Maxillofac Pathol. 2012 May;16(2):183-8. doi: 10.4103/0973-029X.98496. PMID: 22923888; PMCID: PMC3424932.
29. Mahendra A, Shreedhar B, Kamboj M, Singh A, Singh A, Agrawal A, Kumar S, Kabiraj A. Epidermal growth factor receptor protein: a biological marker for oral precancer and cancer. Journal of Dental Surgery. 2014;2014.
30. Condurache Hritcu OM, Botez AE, Olinici DT, Onofrei P, Stoica L, Grecu VB, Toader PM, Gheucă-Solovăstru L, Cotrutz EC. Molecular markers associated with potentially malignant oral lesions (Review). Exp Ther Med. 2021 Aug;22(2):834. doi: 10.3892/etm.2021.10266. Epub 2021 Jun 4. PMID:34149880; PMCID: PMC8200803.
31. Jaiswal PK, Goel A, Mittal RD. Survivin: A molecular biomarker in cancer. Indian Journal of Medical Research. 2015 Apr;141(4):389-397. DOI: 10.4103/0971-5916.159250.
32. Eckert AW, Kappler M, Große I, Wickenhauser C, Seliger B. Current Understanding of the HIF-1-Dependent Metabolism in Oral Squamous Cell Carcinoma. *Int J Mol Sci.* 2020 Aug 24;21(17):6083. doi: 10.3390/ijms21176083. PMID: 32846951; PMCID: PMC7504563.
33. Yang H, Zhong JT, Zhou SH, Han HM. Roles of GLUT-1 and HK-II expression in the biological behavior of head and neck cancer. *Oncotarget.* 2019 Apr 30;10(32):3066-3083. doi:10.18632/oncotarget.24684. PMID: 31105886; PMCID: PMC6508962.
34. Reyes M, Flores T, Betancur D, Peña-Oyarzún D, Torres VA (2020) Wnt/ β -Catenin Signaling in Oral Carcinogenesis. International Journal of Molecular Sciences 21:4682
35. Chaves FN, Bezerra TM, Moraes DC, dos Santos Costa SF, Silva PG, Alves AP, Costa FW, Bernardes VF, Pereira KM. Loss of heterozygosity and immunoexpression of PTEN in oral epithelial dysplasia and squamous cell carcinoma. Experimental and Molecular Pathology. 2020 Feb 1;112:104341.
36. Sawant SS, Vaidya Mm, Chaukar DA, Alam H, Dmello C, Gangadaran P, et al. Clinical significance of aberrant vimentin expression in oral premalignant lesions and carcinomas. Oral Dis. 2014;20:453–65.

37. Ivaska J, Pallari HM, Nevo J, Eriksson JE. Novel functions of vimentin in cell adhesion, migration, and signaling. *Exp Cell Res*. 2007;313:2050–62
38. Akhtar K, Ara A, Siddiqui SA, Sherwani RK. Diagnostic and prognostic significance of E-Cadherin and vimentin in oral cancer metastasis. *Ann Pathol Lab Med*. 2016;03:A8–13.
39. Mishra I, Gaikwad P, Sahu A. Evaluation of e-cadherin and vimentin expression for different grades of oral epithelial dysplasia and oral squamous cell carcinoma - An immunohistochemical study. *J Oral Maxillofac Pathol*. 2022 Apr-Jun;26(2):285-286. doi: 10.4103/jomfp.JOMFP_166_20. Epub 2022 Jun 28. PMID: 35968190; PMCID: PMC9364642.
40. Akhtar K, Ara A, Siddiqui SA, Sherwani RK. Diagnostic and prognostic significance of E-cadherin and vimentin in oral cancer metastasis. *Ann Pathol Lab Med*. 2016;3
41. Akhtar K, Ara A, Siddiqui SA, Sherwani RK. Transition of immunohistochemical expression of e-cadherin and vimentin from premalignant to malignant lesions of oral cavity and oropharynx. *Oman Med J*. 2016;31:165.
42. Sathish II, Asokan K, C L K, Ramanathan A. Expression of E- Cadherin and Levels of Dysplasia in Oral Leukoplakia - A Prospective Cohort Study. *Asian Pac J Cancer Prev*. 2020 Feb 1;21(2):405-410. doi: 10.31557/APJCP.2020.21.2.405. PMID: 32102518; PMCID: PMC7332145.
43. Vogler M. BCL2A1: The underdog in the BCL2 family. *Cell Death Differ*. 2012;19:67–74
44. Krajewski S, Chatten J, Hanada M, Reed JC. Immunohistochemical analysis of the Bcl-2 oncoprotein in human neuroblastomas. Comparisons with tumor cell differentiation and N-Myc protein. *Lab Invest*. 1995;72:42–54
45. Tanda N, Mori S, Saito K, Ikawa K, Sakamoto S. Expression of apoptotic signaling proteins in leukoplakia and oral lichen planus: Quantitative and topographical studies. *J Oral Pathol Med*. 2000;29:385–93
46. Kannan K, Latha PN, Shanmugam G. Expression of bcl-2 oncoprotein in Indian oral squamous cell carcinomas. *Oral Oncol*. 1998;34:373–6
47. Juneja, S; Chaitanya, N Babu1; Agarwal, M2. Immunohistochemical expression of Bcl-2 in oral epithelial dysplasia and oral squamous cell carcinoma. *Indian Journal of Cancer* 52(4):p 505-510, Oct–Dec 2015. | DOI: 10.4103/0019-509X.178411
48. Nasry, Walaa Rodriguez-Lecompte, Juan Martin, Chelsea. (2018). Role of COX-2/PGE2 Mediated Inflammation in Oral Squamous Cell Carcinoma. *Cancers*. 10. 348. 10.3390/cancers10100348.
49. Marconi GD, Della Rocca Y, Fonticoli L, Melfi F, Rajan TS, Carradori S, Pizzicannella J, Trubiani O, Diomedea F. C-Myc Expression in Oral Squamous Cell Carcinoma: Molecular Mechanisms in Cell Survival and Cancer Progression. *Pharmaceuticals (Basel)*. 2022 Jul 19;15(7):890. doi:10.3390/ph15070890. PMID: 35890188; PMCID: PMC9316231.
50. Natesan SC, Ramakrishnan BP, Krishnapillai R, Thomas P. Immunohistochemical Expression of Fascinin in Oral Epithelial Dysplasia and Oral Squamous Cell Carcinoma. *World J Oral Pathol*. 2019 Sep;10(5):341.
51. Folkman J. What is the evidence that tumors are angiogenesis dependent? *J Natl Cancer Inst* 1990;82(1):4–6.

52. Speight PM, Morgan PR. The natural history and pathology of oral cancer and precancer. *CommunDent Health* 1993; 10(Suppl 1):31–41
53. Macluskey M, Chandrachud LM, Pazouki S, Green M, Chisholm DM, Ogden GR, et al. Apoptosis, proliferation, and angiogenesis in oral tissues possible relevance to tumour progression. *J Pathol* 2000;191(4):368–75.
54. Pazouki S, Chisholm DM, Adi MM, Carmichael G, Farquharson GR, Ogden GR, et al. The association between tumour progression and vascularity in the oral mucosa. *J Pathol* 1997;183(1):39–43
55. Penfold CN, Partridge M, Rojas R, Langdon JD. The role of angiogenesis in the spread of oral squamous cell carcinoma. *Br J Oral Maxillofac Surg* 1996;34(1):37–41.
56. Tipoe GL, Jin Y, White FH. The relationship between vascularity and cell proliferation in human normal and pathological lesions of the oral cheek epithelium. *Eur J Cancer B Oral Oncol* 1996;32B(1):24–31.
57. Rajeswari, Parvathy Janardhanan, Mahija Suresh, Rakesh Savithri, Vindhya Aravind, Thara Raveendran, Greeshma. (2020). Expression of CK 19 as a biomarker in early detection of oral squamous cell carcinoma. *Journal of Oral and Maxillofacial Pathology*. 24. 523-529. [10.4103/jomfp.JOMFP_302_19](https://doi.org/10.4103/jomfp.JOMFP_302_19).
58. Lindberg K, Rheinwald JG. Suprabasal 40kd keratin (K19) expression as an immunohistologic marker of premalignancy in oral epithelium. *Am J Pathol* 1989;134:89-98.
59. Frohwitter G, Buerger H, VAN Diest PJ, Korsching E, Kleinheinz J, Fillies T. Cytokeratin and protein expression patterns in squamous cell carcinoma of the oral cavity provide evidence for two distinct pathogenetic pathways. *Oncol Lett* 2016;12:107-13.
60. Yamauchi K, Fujioka Y, Kogashiwa Y, Kohno N. Quantitative expression study of four cytokeratins and p63 in squamous cell carcinoma of the tongue: Suitability for sentinel node navigation surgery using one-step nucleic acid amplification. *J Clin Pathol* 2011;64:875-9.
61. Khanom R, Sakamoto K, Pal SK, Shimada Y, Morita K, Omura K, et al. Expression of basal cell keratin 15 and keratin 19 in oral squamous neoplasms represents diverse pathophysiologies. *Histol Histopathol* 2012;27:949-59
62. Ghazi N, Saghravanian N, Taghi Shakeri M, Jamali M. Evaluation of CD44 and TGF- β Expression in Oral Carcinogenesis. *J Dent (Shiraz)*. 2021 Mar;22(1):33-40.
63. De Vicente, Juan & Molino, Paula & Rodrigo, Juan & Allonca, Eva & Hermida Prado, Francisco & Granda-Díaz, Rocío & Santamarta, Tania & García-Pedrero, Juana. (2019). SOX2 Expression Is an Independent Predictor of Oral Cancer Progression. *Journal of Clinical Medicine*. 8. 1744. [10.3390/jcm8101744](https://doi.org/10.3390/jcm8101744).
64. Luna E, Ealber T, Bezerra Thâmara, Silva P, Cavalcante R, Costa F, Alves A, Chaves F, Pereira KMA. CD133 Role in Oral Carcinogenesis. *Asian Pacific Journal of Cancer Prevention*. 2020;21(9):2501-2506. <https://doi.org/10.31557/APJCP.2020.21.9.2501>.
65. Dai Y, Wu Z, Chen Y, Ye X, Wang C, Zhu H. OCT4's role and mechanism underlying oral squamous cell carcinoma. *J Zhejiang Univ Sci B*. 2023 Sep 15;24(9):796-806. doi:

10.1631/jzus.B2200602.PMID: 37701956; PMCID:
PMC10500100.

66. Grubelnik G, Boštjančič E, Grošelj A, Zidar N. Expression of NANOG and Its Regulation in Oral Squamous Cell Carcinoma. *Biomed Res Int*. 2020 Jul 17;2020:8573793. doi: 10.1155/2020/8573793.PMID: 32733958; PMCID: PMC7383335.
67. Kosmehl H, Berndt A, Strassburger S, Borsi L, Rousselle P, Mandel U, Hyckel P, Zardi L, Katenkamp D. Distribution of laminin and fibronectin isoforms in oral mucosa and oral squamous cell carcinoma. *British journal of cancer*. 1999 Nov;81(6):1071-9.